

ALPN-202, a Conditional CD28 Costimulator and Dual Checkpoint Inhibitor, Utilizes Multiple Mechanisms to Elicit Potent Anti-Tumor Immunity Superior to Checkpoint Blockade

Mark F. Maurer, Chelsea J. Gudgeon, Katherine E. Lewis, Sherri Mudri, Stacey R. Dillon, Martin F. Wolfson, Steven D. Levin, Kristine M. Swiderek, and Stanford L. Peng



Seattle, WA, USA | AlpineImmuneSciences.com | @AlpineImmuneSci

Abstract

INTRODUCTION: Although immune checkpoint inhibitors (CPI) targeting the CTLA-4 and PD-1/PD-L1 pathways have demonstrated significant clinical activity in many cancers, many patients fail to respond, or they develop acquired resistance due to at least in part to insufficient anti-tumor T cell activation and/or exhaustion. Based on this hypothesis, CPIs have been combined with T cell costimulation and shown to enhance and sustain anti-tumor responses preclinically. ALPN-202 is a variant CD80 vlgDTM-Fc fusion protein that mediates PD-L1-dependent CD28 costimulation and inhibits the PD-L1 and CTLA-4 checkpoints (Figure 1). Studies were conducted to characterize the unique mechanism of action of this molecule and further elucidate CD80/PD-L1 biology.

EXPERIMENTAL PROCEDURES: X-ray crystallography and cell-based reporter assays were used to demonstrate the simultaneous binding of PD-L1 and CD28 to the single CD80 IgV domain of ALPN-202. PD-L1-dependent CD28 costimulation was evaluated using primary human T cells stimulated by antigen presenting cells (APC) with or without surface PD-L1. ALPN-202 anti-tumor activity was mechanistically evaluated in vivo using a human PD-L1-transduced MC38 tumor model, by combining ALPN-202 treatment with anti PD-L1, anti CTLA-4, or anti CD28 blocking antibodies. Anti-tumor activity was evaluated by serial tumor volume measurements, and intratumoral immune responses were assessed by flow cytometry and/or RNA-Seq analyses.

DATA SUMMARY: The CD80 vlgD:PD-L1 crystal structure was elucidated at a resolution of 3.15 Å, revealing a non-overlapping binding interface distinct from the previously published CD28:CD80 interaction (Figure 2). Using cell-based reporter and cytokine release assays, monomeric and dimeric CD80 vlgD domains elicited conditional CD28 costimulation which was blocked when combined with anti PD-L1 or anti-CD28 antibodies. In vivo, ALPN-202 demonstrated activity superior to PD-L1 blockade alone, while coadministration with either anti CD28 or anti PD-L1 prevented ALPN-202-mediated conditional CD28 costimulation resulting in reduced anti-tumor activity and RNA signatures consistent with lower T cell infiltration and activation (Figures 3 and 4).

CONCLUSIONS: The CD80 IgV domain utilizes separate, non-competing epitopes to bind CD28 and PD-L1. Exploitation of this attribute via directed evolution-based discovery has yielded ALPN-202, a variant CD80 vlgD-Fc fusion protein capable of simultaneously engaging PD-L1 on tumor cells and CD28 or CTLA-4 on T cells, conferring the unique abilities to block both the PD-L1 and CTLA-4 checkpoints as well as eliciting conditional CD28 costimulation only in the presence of PD-L1. The significantly superior activity of ALPN-202 over CPI-only therapies in vitro and in vivo suggests that conditional CD28 costimulation, together with checkpoint inhibition, may result in meaningfully improved clinical anti-tumor responses.

Results

Figure 1: ALPN-202 is comprised of a variant CD80 IgV domain (vlgDTM) fused to an effectorless IgG Fc

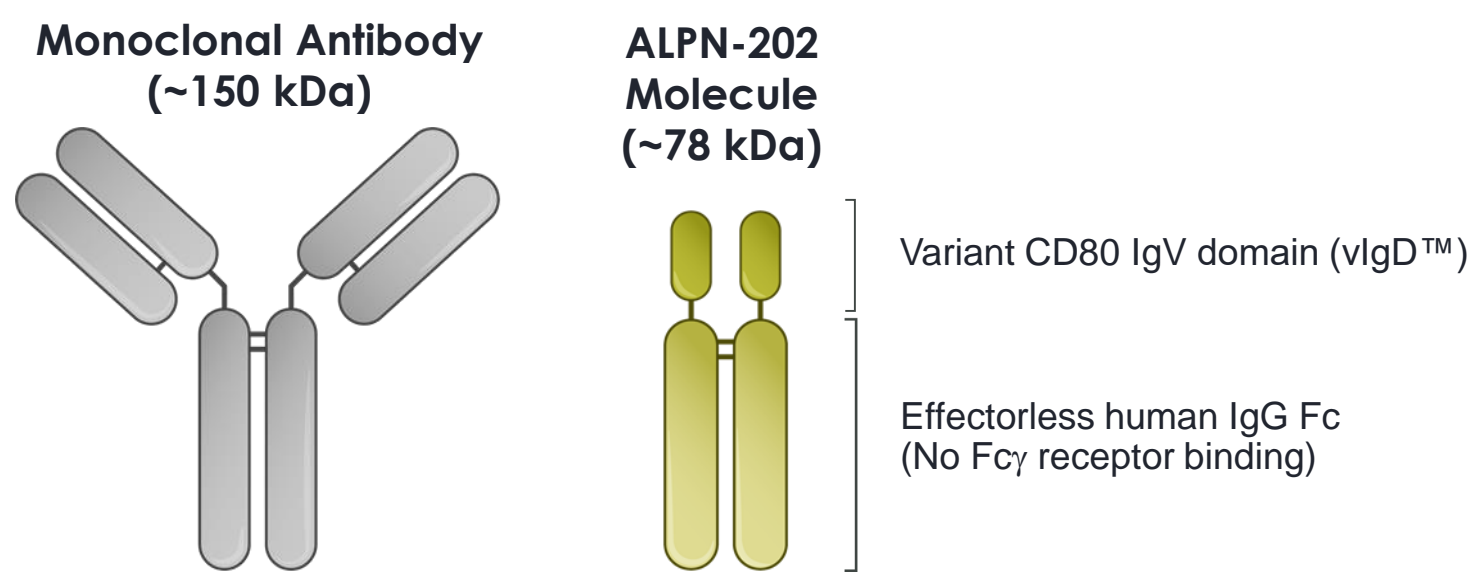
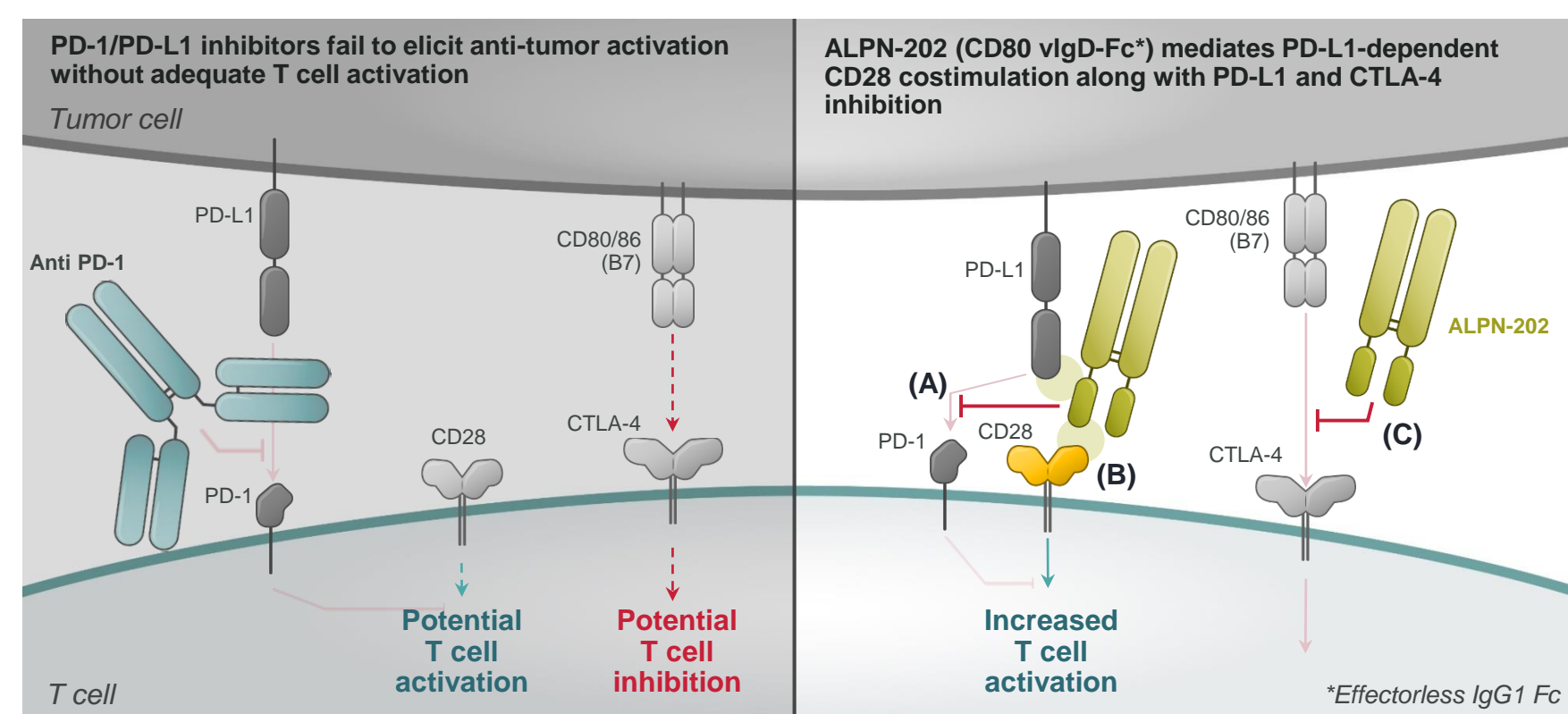
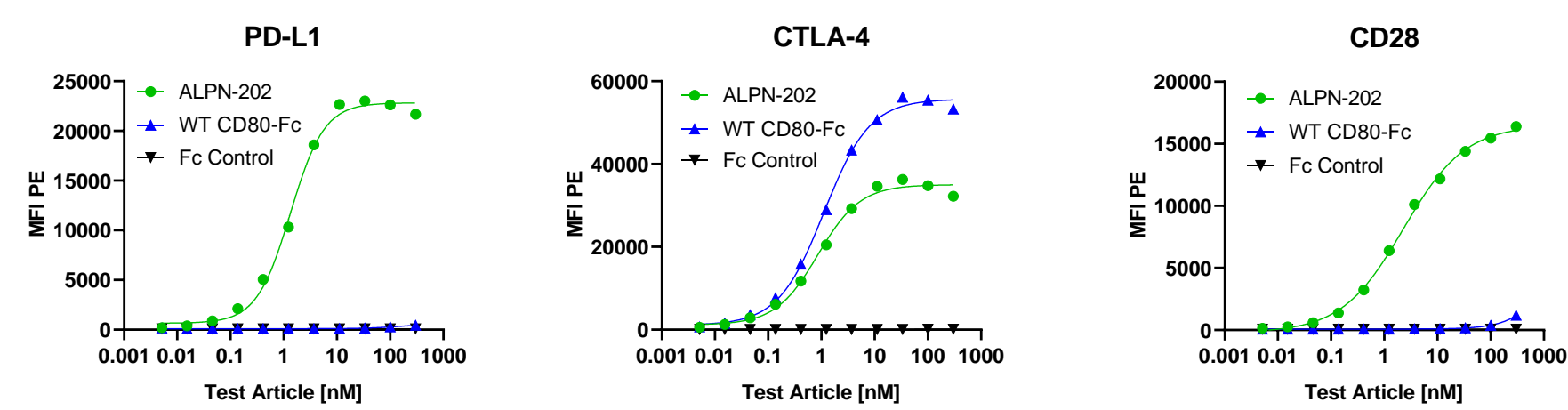


Figure 2: ALPN-202 was engineered for three mechanisms of action: PD-L1 and CTLA-4 antagonism combined with conditional CD28 agonism



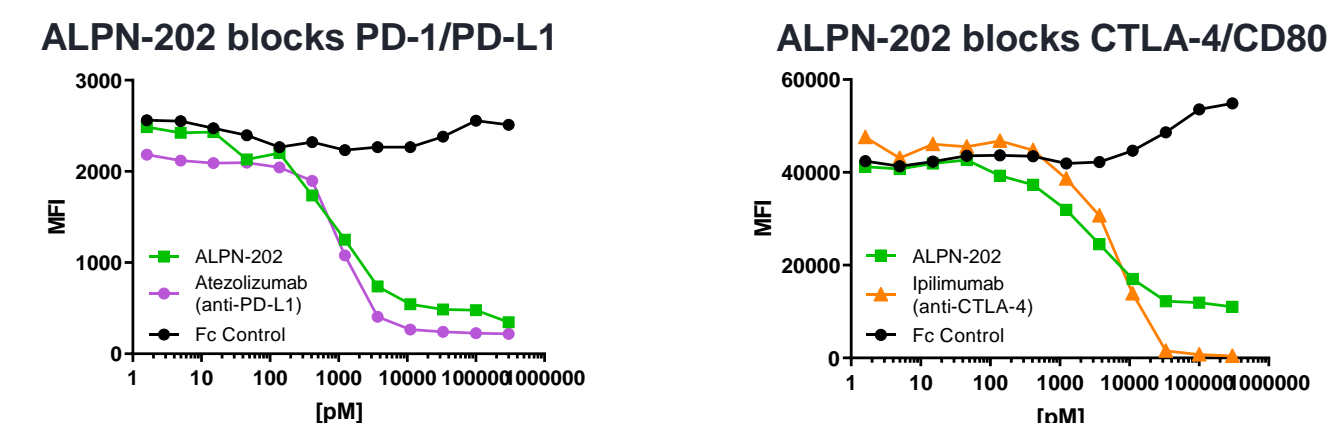
(A) ALPN-202 binds PD-L1 expressed on tumor, blocking PD-L1/PD-1 interactions. (B) Localized, tethered ALPN-202 is able to provide a trans CD28 signal to T cells making contact with the tumor (PD-L1-dependent CD28 costimulation). (C) Additionally, ALPN-202 binds CTLA-4 expressed on T cells, decreasing competition for CD28 signaling, lowering CD3/CD28 signaling thresholds, and promoting TCR repertoire expansion in the periphery.

Figure 3: ALPN-202 has elevated PD-L1 and CD28 affinity relative to wild-type CD80



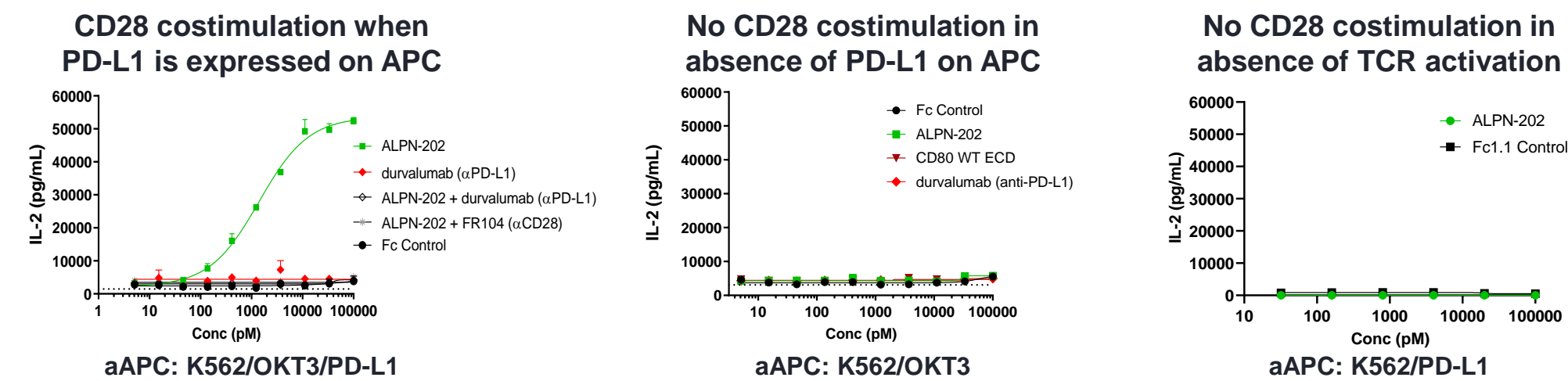
CHO cells stably expressing PD-L1 or CTLA-4, or Jurkat cells endogenously expressing CD28, were used to measure ALPN-202 binding affinity by flow cytometry. ALPN-202 displayed higher affinity for PD-L1 and CD28 than wild-type CD80-Fc.

Figure 4: ALPN-202 antagonizes PD-L1 and CTLA-4



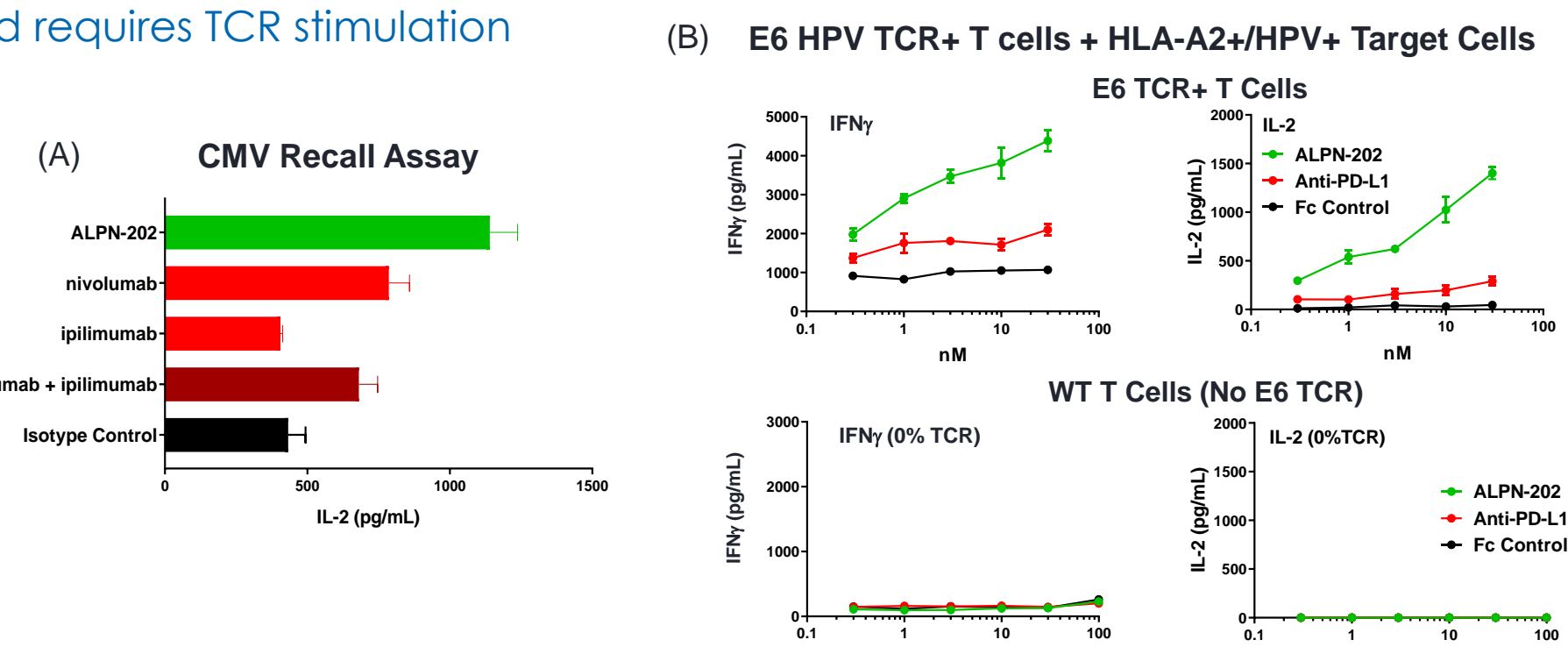
CHO cells stably expressing human PD-L1 (A) or CTLA-4 (B) were plated with titrated ALPN-202 or antibody controls. Cells were washed before adding AF647-conjugated WT PD-1 or CD80-Fc respectively. Binding of conjugated proteins was measured by flow cytometry.

Figure 5: CD28 costimulation requires TCR signal and expression of PD-L1 on the APC



K562 artificial APC (aAPC) stably expressing transmembrane anti-human CD3 and/or human PD-L1 were plated in the presence of primary human T cells and a titration of ALPN-202 or controls. Culture supernatants were harvested at 24 hours and human IL-2 measured by ELISA.

Figure 6: ALPN-202 activity in vitro is more potent than dual checkpoint inhibition and requires TCR stimulation



(A) PBMC were stimulated with CMV lysate in the presence of ALPN-202, anti PD-1 (nivolumab), anti CTLA-4 (ipilimumab), or the combination and IL-2 production measured at 48hr. (B) E6 TCR-transgenic T cells were cultured with the HPV+ tumor line SCC-152. ALPN-202 activity was more potent than anti PD-L1 alone (durvalumab) and dependent on TCR stimulation.

Figure 7: Crystal structure reveals ALPN-202 vlgD binds PD-L1 and CD28/CTLA-4 on opposing surfaces

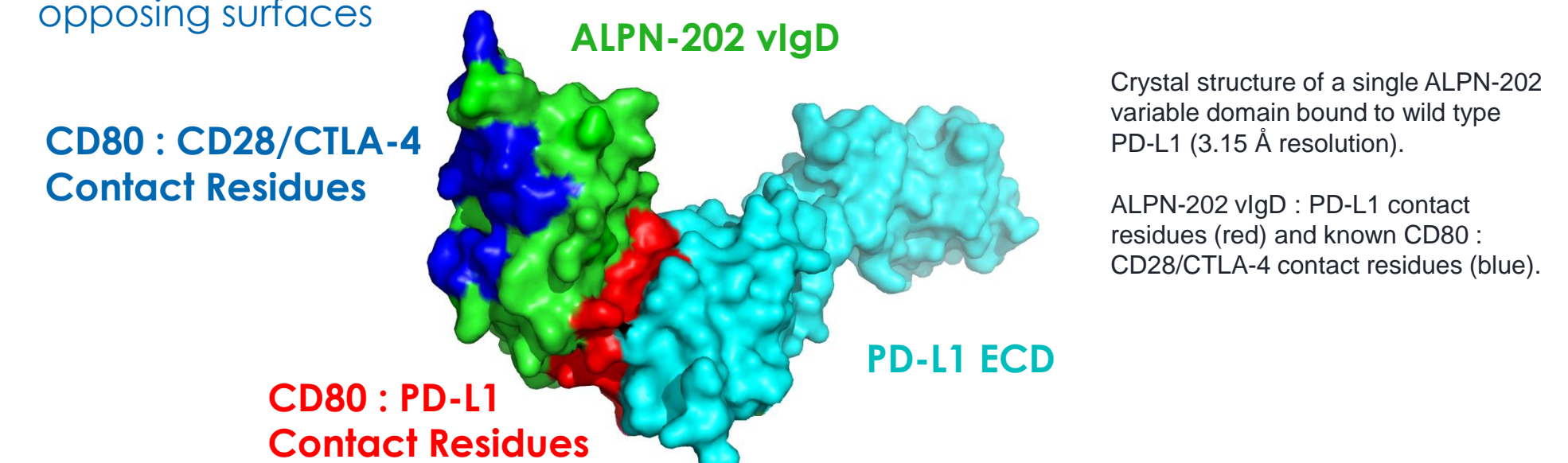
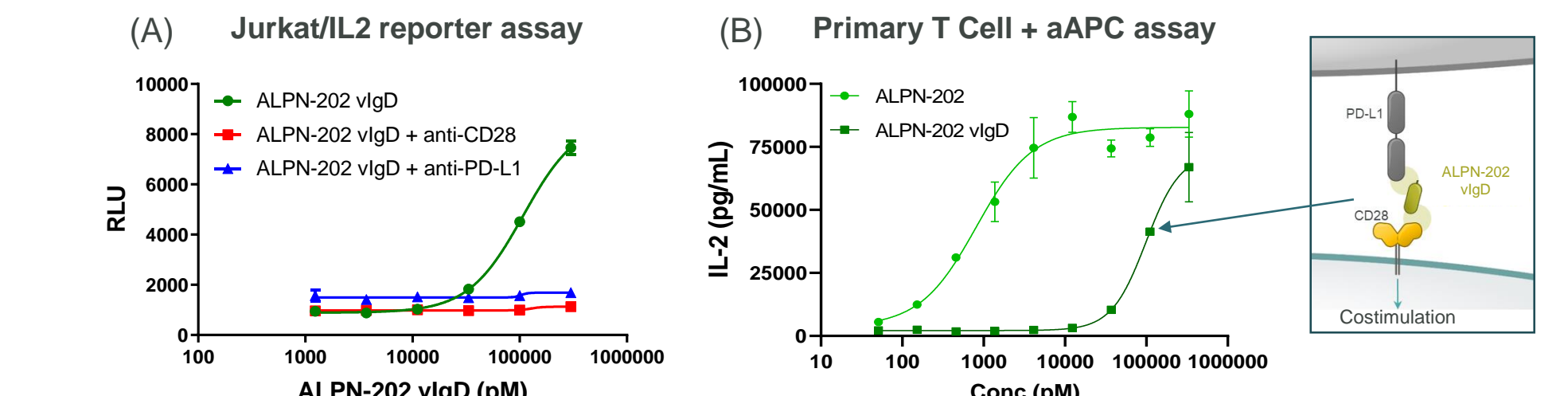
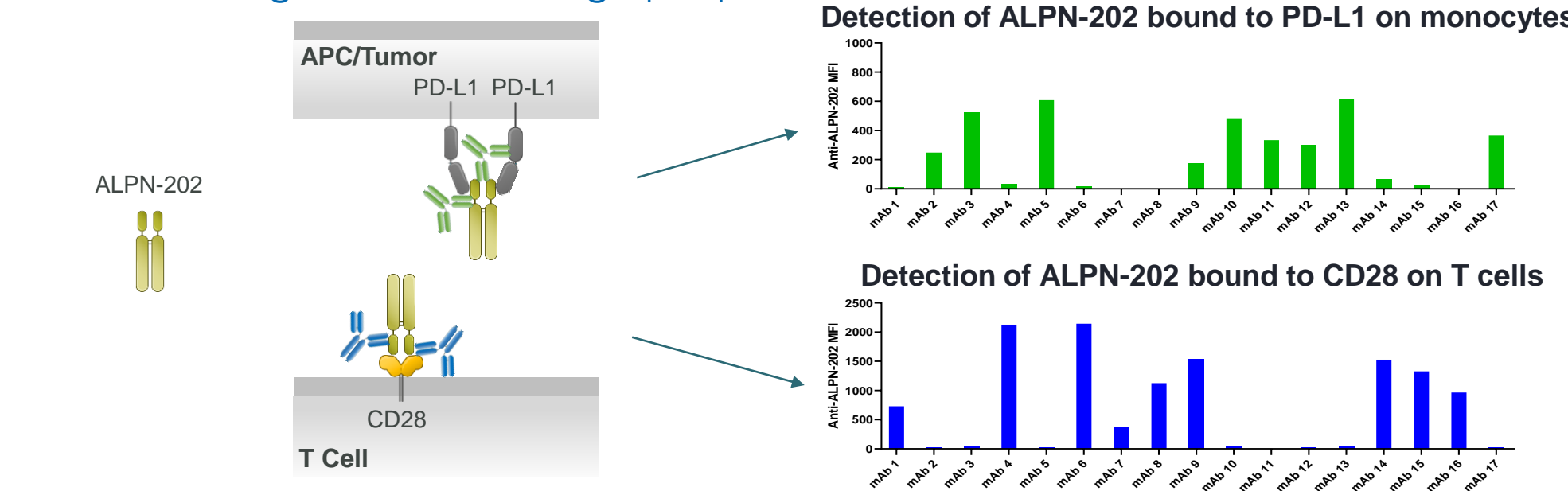


Figure 8: A single ALPN-202 vlgD domain induces PD-L1-dependent CD28 agonism



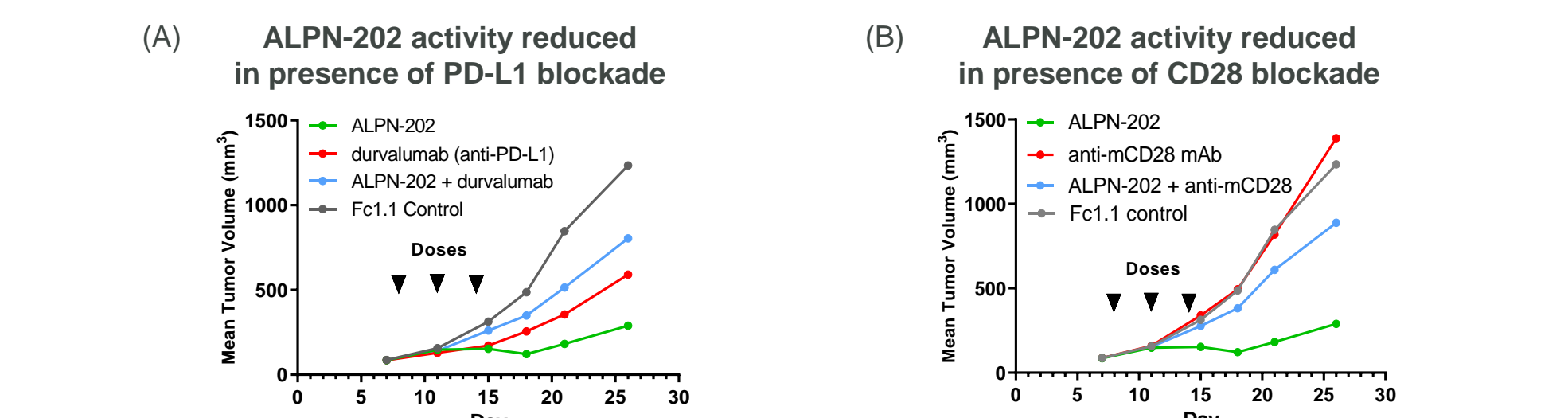
K562 artificial APC (aAPC) expressing anti-human CD3 (OKT3) and WT PD-L1 were plated in the presence of Jurkat/IL2 reporter cells (A) or primary human T cells (B) and incubated with a titration of ALPN-202 alone or with anti CD28 or anti PD-L1 control antibodies. IL-2 reporter activity after 5 hours (RLU) or IL-2 production at 24 hours was measured.

Figure 9: Antibodies can differentiate ALPN-202 bound to CD28 or PD-L1, corroborating different binding epitopes



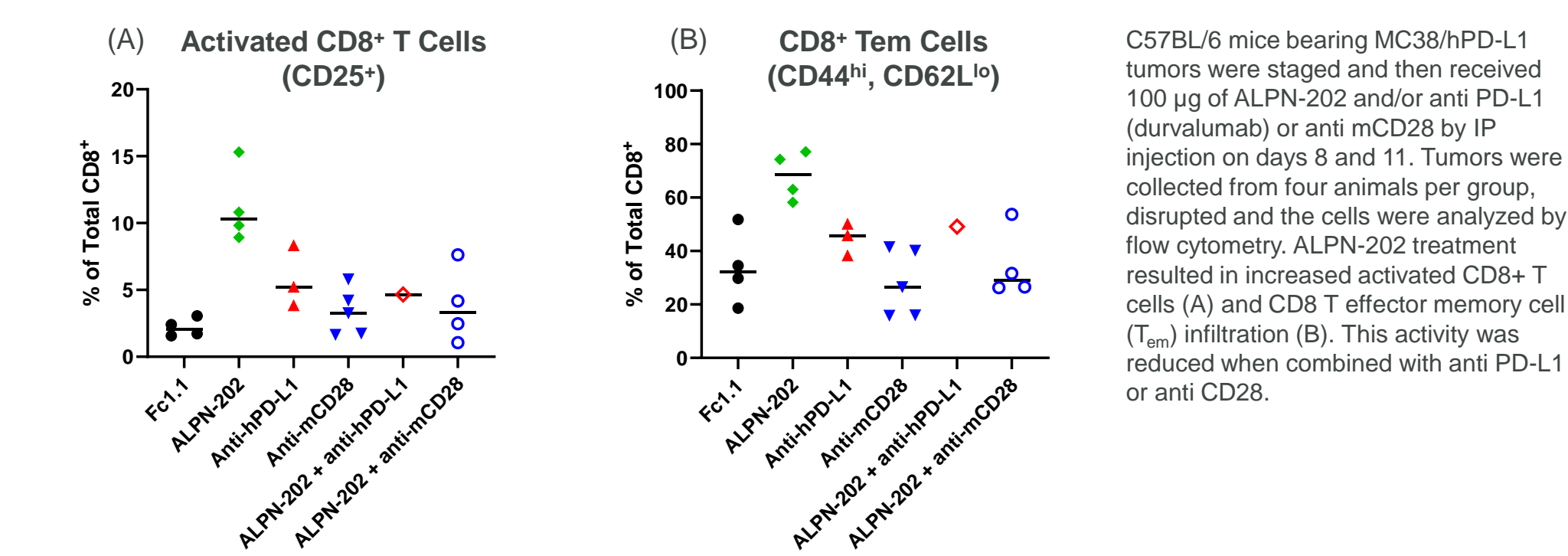
A panel of 17 mouse anti ALPN-202 antibodies were screened for detection of ALPN-202 bound to CD28 on T cells or PD-L1 on activated monocytes. Two distinct groups of antibodies were identified suggesting ALPN-202 binds CD28 on one epitope and PD-L1 on a separate, non-competing epitope.

Figure 10: Selective blockade of ALPN-202 targets in vivo confirms PD-L1-dependent CD28 agonism is a major contributor to anti-tumor activity



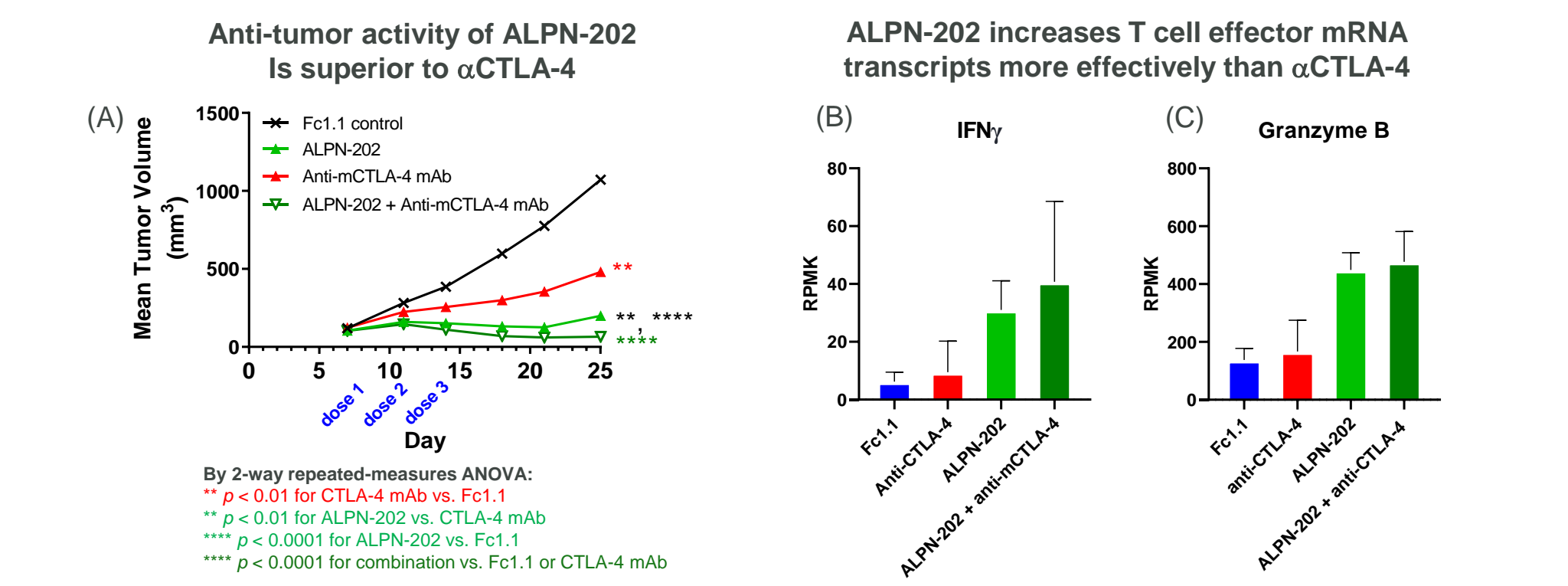
C57BL/6 mice were subcutaneously implanted with human PD-L1 transduced MC38 cells on day 0 and staged on day 7. On days 8, 11, and 14, mice received 100µg of ALPN-202 and/or anti PD-L1 (A) or anti mCD28 (B) antibody by IP injection. Tumor volumes were measured through day 27.

Figure 11: ALPN-202-mediated increase in CD8⁺ T_{em} cell infiltration requires engagement of both CD28 and PD-L1



C57BL/6 mice bearing MC38/hPD-L1 tumors were staged and then received 100 µg of ALPN-202 and/or anti PD-L1 (durvalumab) or anti mCD28 by IP injection on days 8 and 11. Tumors were collected from four animals per group, disrupted and the cells were analyzed by flow cytometry. ALPN-202 treatment resulted in increased activated CD8+ T cells (A) and CD8 T effector memory cell (T_{em}) infiltration (B). This activity was reduced when combined with anti PD-L1 or anti CD28.

Figure 12: ALPN-202 is a potent mono-therapeutic and significantly improves the anti-tumor activity of anti CTLA-4 mAb when given in combination



C57BL/6 mice were implanted s.c. with MC38/hPD-L1 cells on day 0 and staged on day 7. On days 8, 11, and 14, mice received 100 µg of ALPN-202 and/or anti-mouse CTLA-4 antibody (mgG2b) by IP injection. (A) Tumor volumes were measured through day 33. (B, C) 72 hours after second dose, a subset of tumors (n=5/group) was collected, RNA isolated and RNAseq performed. Data as reads per kilobase of transcript, per million mapped reads (RPKM) (mean + SEM).

Visit Poster #467 (K. Lewis, et al): "ALPN-202, a Conditional CD28 Costimulator and Dual Checkpoint Inhibitor, Enhances the Activity of Multiple Standard of Care Modalities"

Summary and Conclusions

- ALPN-202 is a conditional CD28 agonist and a dual PD-L1 and CTLA-4 antagonist
- Blockade of CD28 or PD-L1 inhibits ALPN-202-mediated CD28 costimulation
- ALPN-202 binds PD-L1 and CD28/CTLA-4 at distinct non-overlapping epitopes – enabling its unique functionality
- RNAseq and TIL analyses reveal ALPN-202 treatment results in improved T cell infiltration and pro-inflammatory cytokine production relative to monotherapy CPI
- This potential novel mechanism of action may represent a new, best-in-class immunotherapy for cancer patients
- A first-in-human clinical study with ALPN-202 is in preparation

